Treatment of Metal-Contaminated Wastewater: A comparison of low-cost biosorbents

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Abstract

This study aimed to identify some optimum adsorption conditions for the use of low-cost adsorbent, seaweed (*Ascophyllum nodosum*), sawdust and reed plant (*Phragmites australis*) root, in the treatment of metal contaminated wastewater for the removal of cadmium, chromium and lead. The effect of pH on the absorption capacity of each of these biosorbents was found to be significant and dependent on the metal being removed. Post-adsorption FTIR analysis showed significant binding activities at the nitro N=O groups site in all biosorbents, especially for lead. Competitive metal binding was found to have possibly affected the adsorption capacity for chromium by *A. nodosum* more than it affected sawdust and *P. australis* root. Adsorption is believed to take place mainly by ion exchange particularly at low pH values. *P. australis* root exhibited the highest adsorption for chromium at pH 2, cadmium at pH 10 and lead at pH 7. *A.nodosum* seaweed species demonstrated the highest adsorption capacity of the three biosorbents used in the study, for cadmium at pH 7 and for lead at pH 2. Sawdust proved to be an efficient biosorbent for lead removal only at pH 7 and 10. No significant effect of temperature on adsorption capacity was observed, particularly for cadmium and lead removal.

*Keywords*: Adsorption capacity; heavy metals; pH; reed root; sawdust; seaweed
1. Introduction

Heavy metal contamination of industrial effluents has been an issue of environmental and public health concern since the advent of industrialisation, owing to their non-biodegradable, toxic and bio-accumulative nature (Bailey et al., 1999; Nomanbhay and Palanisamy, 2005). Indicative metals are mostly transition metals like cadmium, copper, zinc, nickel, chromium, as well as other heavy metals such as lead, aluminium, and mercury, amongst others that pose significant risk to soil, water and air environments, their inhabitants and human health via the food chain (Oboh et al., 2009). On discharge of effluents from industries like mining, textile, tannery, metal-plating, petro-chemical, battery and fertilizer production, these metals are deposited in soil, aquatic life-forms and tissues, which form part of the human food-chain. Thus, the toxicity, bio-accumulation and persistence of these metals are transmitted through the food chain and the environment to cause environmental and human health problems (Kanamadi et al., 2006). This situation is most prevalent in tropical developing countries where the enforcement of industrial effluent discharge limits is yet to be accorded the priority it deserves, in contrast to developed countries where stringent environmental quality standards to control pollution by industrial effluents and protect the environment are being adopted and enforced (Dan’Azumi and Bichi, 2010).

Conventional treatment methods have been found to be very expensive and difficult to maintain due to high capital and operational costs as well as extra cost of treating the resultant sludge/secondary waste before disposal as it also poses hazards and pollution risks to the environment (Kumar, 2006). Due to these challenges associated with the conventional methods, growing interest and research into the use of low-cost adsorbents as preferred alternatives to the conventional methods has yielded various successful applications of these low-cost, naturally–occurring and readily available organic adsorbents in the treatment of metal-contaminated industrial effluents using adsorption processes, as reported in various studies (Oboh et al., 2009). The distinct advantages of biosorbents include: low cost, high efficiency, reduced production of chemical or biological sludge, possibility of regeneration of biosorbents and metal recovery (Sud et al., 2008).

Seaweed, sawdust and reed plant root are some of the low-cost readily available organic residues, found naturally in the environment and are considered
waste or by-products. *Ascophyllum nodosum* is considered an effective seaweed for metal adsorption (Romera et al., 2006) and macroscopically produces biosorbent particles, conferring on it a better metal-binding capacity than most organic or inorganic adsorbents (Freitas et al., 2006). Seaweed is reportedly rich in three main polysaccharides - laminaran, fucoidin and alginate, which are rich in anionic carboxylate and sulphate ready for binding at neutral pH (Alluri et al., 2007). The alginate polysaccharide is responsible for its ion-exchange capacity due to its high concentration of carboxyl groups (Mehta and Gaur, 2005; Freitas et al., 2009). The binding surface of seaweed is rich in functional groups like carboxyl, hydroxyl, amine, imidazole, phosphate, sulfhydryl and sulphate groups. However, the carboxyl and sulphate groups are reputedly the most active groups in the binding of metals during adsorption (Romera et al., 2007).

Sawdust, a by-product of the timber industry, which is considered waste except when it is used as packaging material (Vinodhni and Das, 2010) or as cooking fuel, is abundantly available to the point of constituting disposal challenges in timber mills, especially in developing countries. Sawdust consists of three dominant components: cellulose, lignocellulose and lignin, based on which its functional groups reputed to be actively involved in adsorption include C=CH, C=C, C-OH and C-O-C groups (Abdel-Ghani et al., 2007). Its main mechanisms of adsorption are ion exchange and hydrogen binding (Shukla et al., 2002), as well as, chelation and complexation reactions (Asadi et al., 2008).

Reed plant, *Phragmites australis*, is a perennial helophyte grass commonly found in tropical and temperate wetlands where it grows rapidly and exhibits outstanding tolerance of high metal concentrations (Batty, 2003). It is the most commonly employed plant species in the treatment and polishing of wastewater in constructed wetlands where it enhances the break-down organic pollutants by adsorption, accumulation and oxidation via aeration systems around the root zones (Lee and Scholz, 2007). Reed plant contains high concentrations of lignin and cellulose, two components believed to play important roles in its adsorptive capacity for various heavy metals from solution (Southichak et al., 2006). Carboxylic, carboxylate, lignin and lignin ester aromatic C-C groups have been revealed to actively contribute in adsorptive capacity (Southichak et al., 2006), irrespective of
contrasting research reports on its suitability as a biosorbent (Batty, 2003) or otherwise (Lee and Scholz, 2007).

The aim of this study is to identify optimum adsorption conditions for the use of seaweed (*Ascophyllum nodosum*), sawdust and reed plant (*Phragmites australis*) root in the removal of cadmium, chromium and lead from metal-contaminated wastewaters. Cadmium and lead constitute two of the three heavy metals listed by the World Health Organisation (WHO) as the ‘Big Three’ toxic metals, along with Mercury, while chromium was selected because of its ubiquitous presence in various industrial effluents, which are of interest in this study.

2. Materials and methods

2.1 Collection and preliminary treatment of biosorbent samples

The seaweed sample was collected from the beaches of Abroath, Scotland. The reed plant sample was collected from reed-beds in the mini wastewater treatment works serving the Belmont Centre, located in Miegle near Dundee, Scotland and the sawdust sample was collected from the University of Abertay Dundee wood works workshop.

2.2 Preliminary treatment/Preparation of biosorbent samples

The biosorbents were devoid of any chemical pre-treatment or immobilisation and were used in their natural forms after washing and drying. Samples of seaweed (SW) reed (RR) plant and sawdust (SD) used were collected separately. Prior to any preparation, reed plant root was separated from the leaves by cutting. Seaweed and reed plant root were washed separately using tap water, followed by distilled water to remove the sand and stones entangled in the root systems of each sample. Both biosorbent samples were then dried separately in the oven at 105°C for 24 hours. Sawdust was cleaned by hand-picking visible contaminants like wood-splinters and then oven-dried at 105°C for only 3 hours. To optimise the available surface area for the adsorption process, each of the three samples was blended into a powder using a commercial blender and further reduced to finer particles by a 0.5 mm particle-size rotor mill (Fritsch Rotor Speed Mill-Pulverisette 14). Samples were stored separately in clearly-labelled, air-tight, transparent containers at room temperature until required for use.
2.3 Preparation of metal stock standard and dilute solutions

Stock standard solution of each metal was prepared to the strength 1000 mg/l. The procedure for stock standard solution preparation was as follows: Each of 1g of pure cadmium metal (powder form supplied by BDH), 3.735 g of potassium chromate, K\(_2\)CrO\(_4\) and 1.598 g of lead nitrate, Pb(NO\(_3\))\(_3\), was weighed on an Oertling NA 114 scale and put into a separate 100 ml beaker.

Cadmium was dissolved in a mixed solution of 15 ml each of deionised water and concentrated HCl to give a clear solution and heated for 10 minutes. It was allowed to cool and made up to 1 litre solution with 1% (v/v) HCl. Potassium chromate was dissolved in 40 ml of deionised water to give a yellow-coloured solution and diluted to 1 litre with more deionised water. Lead nitrate was dissolved in 40 ml of 1% (v/v) HNO\(_3\). The solution was heated for 2 minutes and allowed to cool at room temperature before being made up to 1 litre with more 1% (v/v) HNO\(_3\). Each solution was then stored in a clearly-labelled 1 litre clear-glass reagent bottle.

2.4 Initial characterisation of biosorbents

2.4.1 Determination of initial metal content of biosorbents

The initial metal content of the biosorbents was determined using the dry ashing and wet digestion methods. This initial metal content determination was deemed necessary in order to determine the actual amount of heavy metals under study present in the biosorbents prior to the adsorption experiments.

2.4.1.1 Dry ashing

Duplicates of 1±0.005 g each of seaweed, reed plant root and sawdust dry powder samples was weighed and placed in porcelain crucibles. The crucibles were then placed in the furnace overnight at 550°C for ashing. The ashed samples were then allowed to cool down at room temperature for approximately 10 minutes and then introduced into 5 ml of 20% HCl to dissolve the residue. The 20% HCl was prepared by adding 20 ml of concentrated HCl to 80 ml of distilled water in a 100 ml conical flask. Each solution was then filtered by suction force through an acid-washed filter paper placed in a filter-flask. The filter paper was then rinsed into the solution in the filter-flask before being transferred into a 50 ml volumetric flask where it was diluted to volume with distilled water before mixing. The liquid samples were then
stored at 4°C in a refrigerator for 10 days before analysis using a Perkin Elmer AAnalyst 200 Atomic Absorption Spectrophotometer (AAS).

2.4.1.2 Wet digestion

10 ml of concentrated HNO$_3$ was added to 1±0.005 g of each dry powder biosorbent sample in a 100 ml beaker and allowed to stand overnight. Each beaker was then heated on a hot-plate until the production of red NO$_2$ fumes ceased. Each beaker was then cooled and 2 ml of 70% HClO$_4$ (Perchloric acid) added. Samples were heated a second time and allowed to evaporate to a much reduced volume before being transferred to a 50 ml volumetric flask and diluted to volume with distilled water. The samples from both digestion processes were then analysed for presence of each of the metals under study using Atomic Absorption Spectrometry (AAS).

2.4.2 Determination of functional group distribution within biosorbent samples using Fourier Transform Infrared Spectroscopy (FTIR)

A pre- and post adsorption determination of the types and distribution of functional groups or bonds present in dry biosorbent samples was carried out by Fourier Transform Infrared Spectroscopy using the Perkin Elmer FT-Infra Red Spectrometer: Spectrum RX1. This determination was to facilitate identification of the functional groups responsible for metal binding on each biosorbent surface (Jin and Bai, 2002). Matching the wavelength of light absorbance to standard bond or functional group types that absorbs light energy at that standard wavelength would achieve the identification.

For the FTIR spectroscopy, Potassium Bromate (KBr) crystals were mixed in a ratio of 8:1 with each dry biosorbent sample and blended into a fine powder, which was used to form uniform KBr glass-like pellets. Automatic Infra-red spectra were then plotted out by the spectrometer. The FTIR spectrometry readings were carried out at specific wavelengths and slits for each metal under study viz: cadmium (228.80 nm and 0.7), chromium (357.87 nm and 2.7/0.8) and lead (217.00 nm and 2.7/1.35).

For all three metals used in this study, readings were done in triplicate. Integration time was 3 seconds, read delay was 5 seconds, unit of measurement was mg/l and calibration equations were all linear through zero. The spectra were then printed out for analysis.
2.5 Adsorption capacity experiment design

The adsorption experiments were carried out in batch mode and in duplicates for each metal and biosorbent.

2.5.1 Factors affecting the adsorption process

The factors affecting the adsorption capacity of the three biosorbents were divided into two groups: constant and variable factors.

2.5.1.1 Constant factors

Constant factors were not varied throughout the adsorption experiments. These included (a) the initial metal concentration, fixed at 200 mg/l, (b) biosorbent dose, 1 ± 0.005 g per sample of metal solution, (c) maximum particle size of 0.5 mm for all biosorbent samples (d) contact time per adsorption experiment, 3 hours. It has been reported that about 75% adsorption is usually attained within 30 - 60 minutes (Souag et al., 2009) while equilibrium is usually attained after about 2 hours of adsorption (Fujiwara et al., 2007).

2.5.1.2 Variable factors

Two temperature levels, 23.5°C and 37°C were used in the study. Both temperatures represent the higher average and maximum temperatures obtainable in most tropical countries. The effect of pH on the adsorption process was studied at four pH values- 2, 5, 7 and 10. Four different solutions containing 200 mg/l of each of three metals used for the study (Cr, Pb and Cd) were prepared for each metal and adjusted to each of the four chosen pH values using concentrated ammonium hydroxide, concentrated HCl or concentrated HNO₃. A Hanna Educational pH Meter HI 208 was used in reading the pH values.

2.5.2 Adsorption experiments

The adsorption experiments were carried out in a batch mode. 1±0.005 g of each biosorbent was weighed into 60 ml plastic centrifuge tubes in three sets of duplicates for the three metals. 50 ml of each 200 mg/l metal solution was added to the tubes containing each biosorbent type and shaken by hand at room temperature for approximately 30 seconds to ensure proper mixing of biosorbents and metals in solution. Tubes containing samples were then allowed to stand for 3 hours but were
shaken again 1.5 hours into the three hours for better biosorbent-metal ion contact. At the end of the 3 hours contact time, the samples were centrifuged for 20 minutes each at a speed of 5400 rpm (rotations per minutes), using an Impact-5 Centrifuge. Centrifugation was followed by the separation of the supernatants from the biosorbents by decantation and filtration using Fisherbrand QL 100 filter paper.

2.5.3 Determination of residual metal concentration in supernatant

The residual or post-adsorption concentration of each metal was determined using the standard method of Direct Air-Acetylene Flame Atomic Adsorption Spectroscopy (AAS). Prior to the AAS analysis, the cadmium and chromium solution filtrates were diluted by a factor of 100 to reduce the concentration to 2 mg/l to meet the linear range for the AAS instrument. Lead was diluted by a factor of 10 down to a concentration of 20 mg/l, within the linear range for AAS measurement of lead. Residual concentration of each metal was then analysed using the specified lamp for each metal and at designated wavelengths. The adsorption capacity, depicted by the amount of each metal adsorbed by the respective biosorbents, was obtained by calculating the difference between the initial amount of metal ions added to the biosorbent and the residual amount of metal ions remaining in the post-centrifugation supernatant after adsorption using the formula: 

\[ Q = \frac{(C_o - C_f)}{V/M} \]

where \( Q \) = Amount of metal adsorbed (mg/g), \( C_o \) = Initial metal concentration in solution (mg/l), \( C_f \) = Final metal concentration in supernatant after adsorption (mg/l), \( V \) = Volume of solution (l) and \( M \) = Mass of the biosorbent (g), which in this experiment was approximately 1 g.

3. Results and discussion

3.1 Initial characterisation of biosorbents

Initial characterisation of biosorbents entailed the pre-adsorption determinations of metal content of biosorbents and functional group distributions within each biosorbent samples.

3.1.1 Determination of initial metal content of biosorbents

Initial metal content determination per biosorbent revealed that of all three metals analysed, chromium had the highest pre-adsorption concentration of between
0.30-0.45 mg per gram of each biosorbent, in the order SW > RR > SD as shown in Table 1.

**Table 1**

Initial metal ion concentration per gram of biosorbent

<table>
<thead>
<tr>
<th>Biosorbent type</th>
<th>Metal ion</th>
<th>Initial metal content determination by:</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Wet digestion (mg/g)</td>
<td>Dry ashing (mg/g)</td>
<td></td>
</tr>
<tr>
<td>Seaweed</td>
<td>Cd</td>
<td>0.02</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Cr</td>
<td>0.34</td>
<td>0.45</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Pb</td>
<td>0.01</td>
<td>0.01</td>
<td></td>
</tr>
<tr>
<td>Sawdust</td>
<td>Cd</td>
<td>0.00</td>
<td>0.12</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Cr</td>
<td>0.30</td>
<td>0.32</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Pb</td>
<td>0.00</td>
<td>0.02</td>
<td></td>
</tr>
<tr>
<td>Reed Root</td>
<td>Cd</td>
<td>0.00</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Cr</td>
<td>0.31</td>
<td>0.33</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Pb</td>
<td>0.03</td>
<td>0.00</td>
<td></td>
</tr>
</tbody>
</table>

The relatively high pre-adsorption concentration of chromium in all three biosorbent samples may affect its adsorption capacity.

3.1.2 Determination of functional group distribution within biosorbent samples

Pre-adsorption FTIR results indicate strong presence of such groups as the carboxylic acid or alcoholic O-H bond stretching which may overlap with amine (N-H) bond stretching at peaks between 3250-3400 cm\(^{-1}\) for all three biosorbents. Possible C-H bond stretchings of alkanes at peaks just below 3000 cm\(^{-1}\); C=O bond of carbonyl or amide groups within 1640-1670 cm\(^{-1}\); C-O and O-H bond stretchings of alcohol and ethers at 1000-1260 cm\(^{-1}\) of the finger-print region; the aromatic C-H bond bends at 690-900 cm\(^{-1}\) and possible sulphate or sulfoxide S=O bond around 1050 cm\(^{-1}\) were also observed for only sawdust and reed plant root. Comparison of pre and post-adsorption FTIR results showed only slight changes in the pre-adsorption results, suggesting probable uniformity in manner of adsorption, as all three adsorbents are organic in nature and may bind metals in similar manners.
For the seaweed, no significant change was observed after chromium adsorption with seaweed, which may possibly be due to competitive binding by high pre-adsorption chromium ions. Lead adsorption showed a strikingly deeper trough at 1384.5 cm$^{-1}$ indicating possible binding to the nitro N=O stretching. With sawdust, all three metals caused slightly deeper troughs at 1033 cm$^{-1}$ and 1059 cm$^{-1}$ suggesting metal binding to amine C=N bonds. Changes were observed at 1508.3 cm$^{-1}$ and 1384.4 cm$^{-1}$ for chromium and lead respectively, suggesting binding to nitro N=O bonds. With reed plant (P. australis) root, the major change observed was at 1384.4 cm$^{-1}$ with lead adsorption, suggesting binding to nitro N=O bonds. These identified regions may be indicative of functional groups responsible for the individual metal-binding activity of each biosorbent, apart from the groups indicated in previous studies (Kanamadi et al., 2006).

3.2 Adsorption capacity of absorbents

The results stated were mean absorbent values of duplicated experiments.

3.2.1 Effect of temperature

Experiments carried out with temperature being the sole variable, showed that for a given adsorbent, metal ion and pH value used in the study, there was no significant difference between adsorption capacities at 23.5°C and at 37°C.

3.2.2 Effect of initial pH

Figures 1, 2 and 3 show the variation of adsorption capacity with pH for the three biosorbents. Adsorption of cadmium was observed to increase with increasing pH, being highest at pH 10 for all three biosorbents studied as shown in Fig. 1. Adsorption of chromium was observed to generally decrease with increasing pH with its highest adsorption occurring at pH 2 and no adsorption occurring at pH 10 as shown in Fig. 2. At pH 2, reed plant root exhibited the highest adsorption capacity indicating its highest removal efficiency in acidic solutions. No adsorption for chromium by seaweed was observed throughout the pH range while chromium adsorption by sawdust was significant only at pH 2. The relatively high pre-adsorption concentration of chromium in all three biosorbent samples, shown in Table 1, might have affected adsorption capacity of chromium ions in solution due to competitive binding at the biosorbent chromium-binding sites.
Figure 3 shows that with lead, adsorption capacity decreased with increasing pH for seaweed, increased with increasing pH for sawdust and increased to maximum adsorption at pH 7 for reed plant root before decreasing in alkaline solutions. For sawdust, lead adsorption increased rapidly and steadily with increase in pH, suggesting increasing capacity for lead uptake from acidic to alkaline solutions, with a maximum adsorption capacity of 183.33 mg/g of lead at pH 10. For reed plant root, a consistently high rate of lead uptake was observed throughout the pH range, suggesting the biosorbent’s high efficiency in lead removal from both acidic and alkaline solutions and this characteristic may qualify reed plant root as a biosorbent of choice for various types of industrial effluents as most effluents generally do not have singular pH values.

Figure 1. Effect of pH on cadmium ion adsorption using seaweed (SW-Cd), sawdust (SD-Cd) and reed plant root (RR-Cd)
Figure 2. Effect of pH on chromium adsorption using seaweed (SW-Cr), sawdust (SD-Cr) and reed plant root (RR-Cr)

Figure 3. Effect of pH on lead adsorption using seaweed (SW-Pb), sawdust (SD-Pb) and reed plant root (RR-Pb)
4. Discussion

Figure 4 summarises the adsorptive capacities of each biosorbent for the metals used in the study.

![Bar chart showing adsorption capacities of seaweed (SW), sawdust (SD) and reed plant root (RR) for cadmium (Cd), chromium (Cr) and lead (Pb)]

For cadmium adsorption, seaweed exhibited the most consistent adsorption throughout the pH range. However, its highest cadmium removal of 106.26 mg at pH 10 is far less than the 168.56 mg removed by reed plant root at the same pH, suggesting that reed plant root may possess superior cadmium removal efficiency in alkaline solutions. Conversely, seaweed removal of 86.16 mg of cadmium at pH 7 exceeds the 61.70 mg removed by reed plant root and that may be indicative of seaweed’s better capacity for cadmium adsorption at or near neutral pH values. Sawdust exhibited a generally low adsorptive capacity for cadmium throughout the study. Hence, it appears to be the least efficient biosorbent for cadmium in this study.
Chromium adsorption of 89.20 mg by reed plant root and 30.46 mg by sawdust at pH 2 suggests that, of the three biosorbents studied, reed plant root demonstrated the highest affinity for chromium ions in acidic solutions, followed by sawdust, which appears to lack efficiency in chromium removal throughout the rest of the studied pH range. Seaweed demonstrated no chromium adsorption capacity throughout this study, and as such, may be considered the least efficient in chromium removal from solution.

Lead removal from solution was consistently high for all three biosorbents studied. Reed plant root exhibited the most consistent lead removal cumulatively with highest removal of 183.06 mg occurring at pH 7, suggesting the highest lead removal efficiency of reed plant root may be in neutral or near-neutral pH effluents. Lead adsorption by seaweed remained relatively high in acidic pH values with highest adsorption of 186.82 mg at pH 2, indicating optimal lead removal efficiency of seaweed in acidic solutions. However, lead adsorption by sawdust was only high at pH 7 and 10 with its highest removal of 186.32 mg occurring at pH 10, suggesting optimal lead removal efficiency for sawdust in alkaline effluents.

A central deduction emerging from the above trends in efficiency is that biosorbent efficiency in metal ion removal varies with wastewater effluent pH. This is because metal binding would depend on the availability, types and concentrations of functional groups on the biosorbent surface at different pH values. However, in considering the practical application of biosorbents in industrial wastewater treatment in relation to pH, extreme acidic or alkaline pH values are not considered desirable and environmentally-safe and as such, near-neutral or neutral pH is considered the most suitable as it approximates the pH of most oceans and seas into which treated wastewater is discharged. It would also entail the least health and safety requirements, little or no chemicals for further treatment of the effluent after metal adsorption before discharge and would not alter the pH of the receiving water environment. However, care must be taken in deciding the optimal pH for a biosorbent as the final pH of the effluent after adsorption may vary from the initial pH, depending on the amount of H⁺ and OH⁻ ions released during the adsorption process. Table 2 shows the biosorbent with the highest adsorptive capacity for each metal ion at each pH value.
Table 2

<table>
<thead>
<tr>
<th>Metals</th>
<th>pH 2</th>
<th>pH 5</th>
<th>pH 7</th>
<th>pH 10</th>
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<tr>
<td>Cadmium</td>
<td>SW</td>
<td>SW</td>
<td>SW</td>
<td>RR</td>
</tr>
<tr>
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<td>RR</td>
<td>RR</td>
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</tr>
<tr>
<td>Lead</td>
<td>SW</td>
<td>SW</td>
<td>RR</td>
<td>SD</td>
</tr>
</tbody>
</table>

5. Conclusion and perspectives

Results from this study have shown that some low-cost biosorbents may be suitable for the treatment of some metal contaminated wastewaters under certain conditions, with pH playing a more significant role temperature on treatment efficiency. Competitive metal binding was found to significantly affect seaweed’s (A. nodosum) adsorption capacity for chromium than it affected sawdust and reed plant (P. australis) root. Adsorption was found to occur throughout pH range but especially at low pH values. Post-adsorption functional group distribution analysis indicates significant binding activities at the site of the nitro N=O groups in all biosorbents, especially for lead. Reed plant root was found to be most efficient biosorbent for chromium at pH 2, for cadmium at pH 10 and for lead at pH 7. Seaweed yielded the highest adsorption capacity for cadmium at pH 7 and for lead at pH 2. Sawdust proved to be an effective biosorbent for lead removal only at pH 7 and 10. This study indicates that waste water treatment in developing countries could be assisted by low cost biosorbents, with wastewater pH being more important than temperature for the adsorbents and metals used in this study. Further study could consider the most appropriate combination of biosorbents and pretreatment requirements for specific industrial waste waters applications.

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